

## Identification of antimicrobial constituents in essential oil from *Paulownia fortunei* flowers

\*Malik F. H. Ferdosi<sup>1</sup>, Iqra Haider Khan<sup>1</sup>, Arshad Javaid<sup>1</sup>, Tayyaba Sattar<sup>2</sup>, Ayesha Munir<sup>3</sup>

<sup>1</sup>Institute of Agricultural Sciences, University of the Punjab, Quaid-i-Azam Campus, Lahore 54590, Pakistan

<sup>2</sup>Department of Chemistry, Lahore College for Women University, Jail Road Lahore, Pakistan

<sup>3</sup>Institute of Chemistry, University of the Punjab, Quaid-i-Azam Campus, Lahore 54590, Pakistan

\*Corresponding author's email: [fiaz.iags@pu.edu.pk](mailto:fiaz.iags@pu.edu.pk), [malikferdosi@yahoo.com](mailto:malikferdosi@yahoo.com)

### Abstract

The aim of this study was to analyze essential oil from flowers of *Paulownia fortunei* through GC-MS for identification of possible antimicrobial constituents. The essential oil from flowers of *P. fortunei* was extracted by microwave assisted technique. The oil was analyzed by GC-MS that showed the presence of six compounds. Nerolidol was the principal compound in the extract with 82.81% peak area. Other compounds included pentacosane (3.95%), octadecane (4.77%), 5,9-undecadien-2-one, 6,10-dimethyl-, (E)- (3.02%), heptacosane (2.81%), and (E,E,E)-3,7,11,15-tetramethylhexadeca-1,3,6,10,14-pentaene (2.68%). Literature survey showed that the principal compound nerolidol possesses potent antibacterial and antifungal properties.

**Keywords:** Antimicrobial, Essential oil, Flower, GC-MS analysis, *Paulownia fortunei*.

### Introduction

Essential oils (EOs) are natural and volatile compounds extracted as secondary metabolites from various aromatic plants (Perczak *et al.*, 2019). These are insoluble in water and soluble in ether and alcohol. These are generally liquid at room temperature and colorless, often have a distinctive taste as well as pleasant odor (Sarkic and Stappen, 2018). These are a complex mixture of low molecular weight phytoconstituents extracted by steam distillation, dry distillation, cold pressing or solvent extraction (Khan and Dwivedi, 2018). These may constitute 20–200 different plant secondary metabolites belonging to a variety of chemical classes including phenylpropanoids, sesquiterpenes, monoterpenes, terpenoids, aliphatic and few aromatic compounds (Aziz *et al.*, 2018). Plant EOs possess various applications mainly in agriculture, food industries, health and cosmetics (Reddy, 2019; Sharmeen *et al.*, 2021). These can be obtained from different plant organs such as flowers, buds, twigs, leaves, stem, fruits, bark, roots and seeds (Ghaffari *et al.*, 2019). Several researchers have globally screened EOs as a potential source of novel compounds, food preservatives and also to treat infectious diseases (Singh *et al.*, 2021). These exhibit biological properties including antioxidant, anticancer, antimutagenic, antiviral, antifungal, antibacterial, immunomodulatory, anti-inflammatory and antiprotozoal activities (Tariq *et al.*, 2019; Valdivieso-Ugarte *et al.*, 2019).

Dragon tree [*Paulownia fortunei* (Seem.)

Hemsl.], native to China and Taiwan, is a fast-growing perennial tree of family Scrophulariaceae. Its wood is highly suitable for paper production (Kiaei, 2013), with a fiber length of 1.42 mm (Rai *et al.*, 2000). Epicarp of its fruits contain antimicrobial activity against *Bacillus subtilis* and *Staphylococcus aureus* (Cercós, 1982). *P. fortunei* also possesses various medicinal properties. Its seeds are useful to treat diabetes and leaves show marked antioxidant property (He *et al.*, 2016). *Paulownia* spp. were introduced in Pakistan from 1989–95 at 13 locations in Kashmir, Punjab and Khyber Pakhtunkhwa by Pakistan Forest Institute and the seeds were obtained from Chinese Academy of Forestry (Siddiqui and Khan, 1989; Bajwa and Gul, 2000). Different plant species are known to possess a variety of antimicrobial compounds (Banaras *et al.*, 2020; Khan and Javaid, 2020). However, there is not any such report from flowers of *P. fortunei* growing in Pakistan. Therefore, in the present study, GC-MS analysis of essential oil of flowers of *P. fortunei*, collected from Lahore, Pakistan, was carried out for identification of antimicrobial compounds.

### Materials and Methods

Few years back, seedlings of *P. fortunei* were brought from China and were sown at Faculty of Agricultural Sciences, University of the Punjab Lahore Pakistan. The mature fresh flowers were collected during spring 2021 from these plants. The flowers were kept in paper boxes and shifted to the laboratory for the essential oil extraction process.

Microwave assisted procedure was used to extract essential oil from *P. fortunei* flowers at Institute of Chemistry, University of the Punjab Lahore Pakistan. A modified microwave oven was utilized in the experiment, along with glass bends and a condenser. An electric grinder was used to crush 305 g of *P. fortunei* flowers into a paste. The sample was moved to a one-liter round-bottom flask and stabilized within the oven with bends, one of which was connected with a condenser using plastic clamps. The condenser is connected to the collecting flask by another bend. The oven's power level was set to 60 °C for 60 minutes, and the extraction process was initiated. After every 10 minutes, the orientation of the sample-containing flask was manually turned to ensure uniform heating. The essential oil-containing hydrosol from the collecting flask was transferred to the separating funnel and allowed to cool to room temperature. Dimethyl sulphoxide (5 mL) was added to extract the essential oil from the hydrosol. Oil containing DMSO layer sank down to the bottom and collected in screw vial.

The GC-MS analysis was carried out on a Gas Chromatograph (GC) machine model 7890B and that of Mass Spectroscopy (MS) machine model 5977A branded by Agilent Technologies. The column used was DB 5 MS, (30 m × 0.25 µm × 0.25 µm). Injection volume was 1 µL and carrier gas was helium. Oven ramping; initial temperature was 80 °C and then raised 10 °C per minute up to 300 °C. Inlet temperature was 280 °C. MS conditions were as: the source temperature was 230 °C and quadrupole temperature was 150 °C. Chemical compounds were identified by comparison of their spectra with NIST 2017 library and arranged in the ascending order of their retention times. The relative abundance was reported by using their peak areas.

## Results and Discussion

There were six compounds in the methanolic extract of *P. fortunei* as shown in Fig. 1 and Table 1. Nerolidol was the principal constituent in this showing 82.81% peak area. Other compounds present in the extract were pentacosane (3.95%), octadecane (4.77%), 5,9-undecadien-2-one, 6,10-dimethyl-, (E)- (3.02%), heptacosane (2.81%), and (E,E,E)-3,7,11,15-tetramethylhexadeca-1,3,6,10,14-pentaene (2.68%). Mass spectra and structures of these compounds are given in Fig. 2. Essential oil of *P. tomentosa* flower from Egypt contained geranyl geraniol (18.05%) as the predominant compound followed by hexatriacontane (11.61%) with antibacterial activity against *Bacillus subtilis*, *Escherichia coli* and *Staphylococcus aureus* (Ibrahim *et al.*, 2013).

Nerolidol, the major compound in the extract is known to have marked antimicrobial potential. It is a sesquiterpene alcohol and is found in essential oils of many plant species including *Citrus aurantium*, *Hedychium coccineum* and *Jasminum*

*grandiflorum* (Gurib-Fakim *et al.*, 2002; Jirovetz *et al.*, 2007; Ammar *et al.*, 2012). This compound is a flavoring agent, and is also used in as a fragrance agent in detergents and cosmetics (Lapczynski *et al.*, 2008). Nerolidol and its various derivatives namely (-)- $\alpha$ -bisabolol, *cis*-nerolidol, *O*-ethyl-nerolidol, and *O*-methyl-nerolidol are known for their antibacterial and antifungal activities (Krist *et al.*, 2015). Nerolidol isolated from essential oil of *Chamaecyparis obtusa* showed antifungal activity against *Microsporum gypseum* (Lee *et al.*, 2007). In addition to its own antibacterial potential, this compound also enhances action of other antibiotic drug by increasing bacterial permeability (Brehm-Stecher and Johnson, 2003). It disrupts barrier function of bacterial membrane by leaking potassium ion from *Staphylococcus aureus* (Inoue *et al.*, 2004).

Octadecane, an alkane, was the second most abundant compound with 4.77% peak area. It has been identified as a highly abundant compound in bulb oil of *Aliumnigrum* (30.5%) (Rouis-Soussi *et al.*, 2014). It has also been identified in many other plant species including *Rhaponticum acaule* and *Trichosanthes dioica* (Boussaada *et al.*, 2008; Khatua *et al.*, 2016). Similarly, two other compounds identified in the present study namely pentacosane and heptacosane were also alkanes. Presence of alkanes as volatile compounds in many plants species has been reported in many publications (Adeleye *et al.*, 2011). Marrufo *et al.* (2013) reported that hydrocarbons showed 91.1% of oils of *Moringaoleifera*. The major compounds were pentacosane (13.3%), hexacosane (13.9%), and heptacosane (11.4%). The oil was found having antibacterial and antifungal activities mainly due to the presence of these hydrocarbons. He (2009) reported that some alkanes are known for having marked antimicrobial effect particularly against *Escherichia coli* and *Staphylococcus aureus*.

5,9-Undecadien-2-one, 6,10-dimethyl-, (E)-, also known as geranylacetone, is a colorless oil. It is a flavor component of various plant species such as tomato, mango and rice (Pino *et al.*, 2005). It has also been found as a minor compounds in oil of an endophytic fungus *Gliomastix murorum* isolated from *Paris polyphylla* var. *yunnanensis* (Zhao *et al.*, 2009). It has trypanostatic activity and can protect animals from *Trypanosoma congolense*-induced anaemia by inhibiting sialidase (Saad *et al.*, 2019).

(E,E,E)-3,7,11,15-Tetramethylhexadeca-1,3,6,10,14-pentaene, also known as  $\alpha$ -springene, is a diterpene. Previously, it was identified in *Ligularia fischeri* var. *spiciformis* with strong cytotoxicity against HL-60 (Lee *et al.*, 2002). This compound was also found in essential oils of *Murraya exotica* and *Teucriummarum* as a major component (Raina *et al.*, 2006; Djabou *et al.*, 2013).

## Conclusion

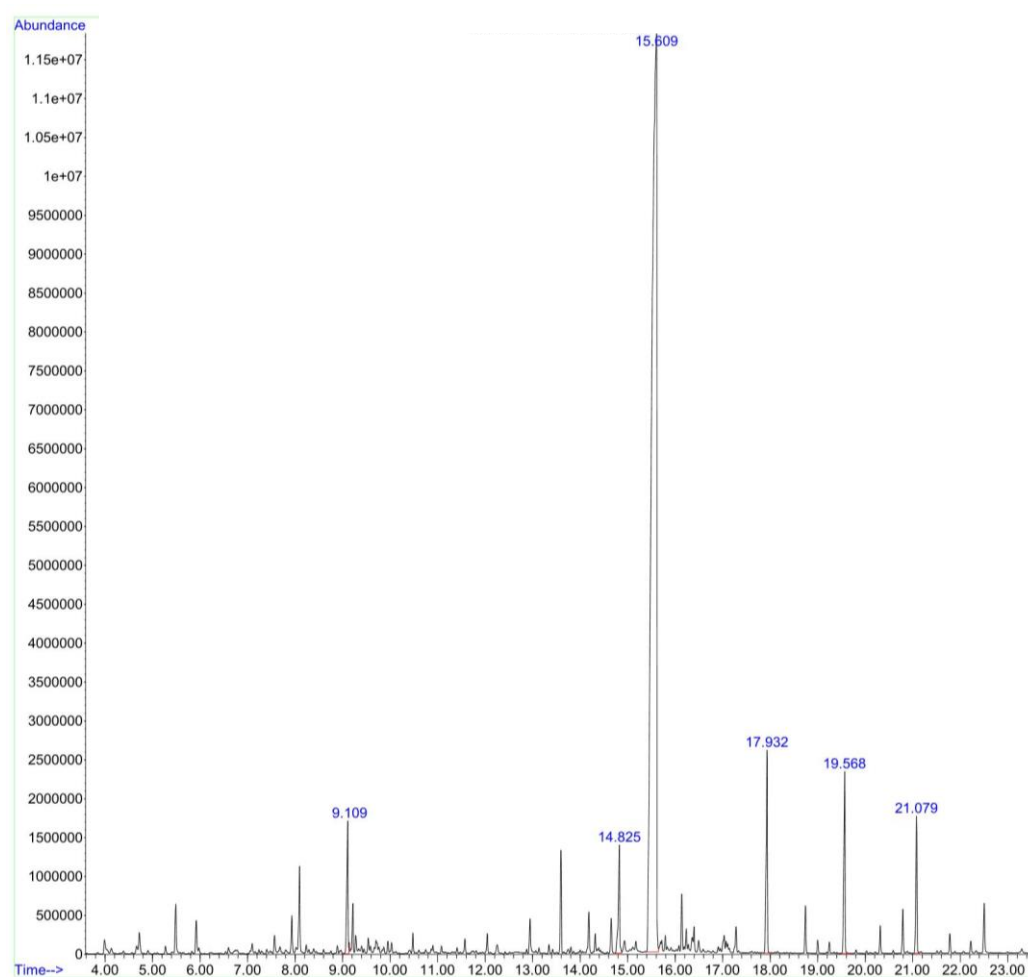
The main component in essential oil of *P. fortunei* was nerolidol that is known for its antibacterial and antifungal activities.

## Acknowledgement

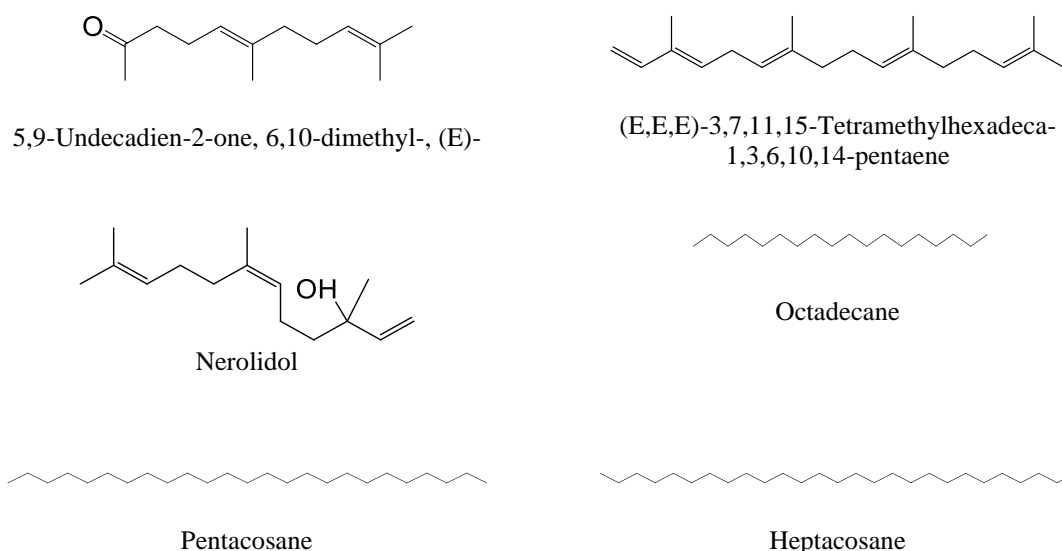
Dr. Muhammad Ashfaq brought seedlings of *P. fortunei* from China and cultivated at Institute of Agricultural Sciences, Punjab University Lahore.

**Table 1:** List of compounds in methanolic flower extract of *Paulownia fortunei* identified by GC-MS analysis.

Sr. No.	Names of compounds	Molecular formula	Molecular weight	Retention time (min)	Peak area (%)
1	5,9-Undecadien-2-one, 6,10-dimethyl-, (E)-	C <sub>13</sub> H <sub>22</sub> O	194.31	9.109	3.02
2	(E,E,E)-3,7,11,15-Tetramethylhexadeca-1,3,6,10,14-pentaene	C <sub>20</sub> H <sub>32</sub>	272.46	14.825	2.68
3	Nerolidol	C <sub>15</sub> H <sub>26</sub> O	222.37	15.609	82.81
4	Octadecane	C <sub>18</sub> H <sub>38</sub>	254.49	17.932	4.77
5	Pentacosane	C <sub>25</sub> H <sub>52</sub>	352.68	19.568	3.91
6	Heptacosane	C <sub>27</sub> H <sub>56</sub>	383.73	21.079	2.81



**Fig. 1:** GC-MS chromatogram of methanolic flower extract of *Paulownia fortunei*.



**Fig. 2:** Structures of compounds identified in methanolic flower extract of *Paulownia fortunei*.

## References

- Adeleye IA, Daniels FV, Omadime M, 2011. Characterization of volatile components of Epa-Ijebu: A native wonder cure recipe. *J. Pharmacol. Toxicol.*, **6**: 97-100.
- Ammar AH, Bouajila J, Lebrihi A, Mathieu F, 2012. Chemical composition and *in vitro* antimicrobial and antioxidant activities of *Citrus aurantium* L. flowers essential oil (neroli oil). *Pak. J. Biol. Sci.*, **15**: 1034-1040.
- Aziz ZA, Ahmad A, Setapar SHM, Karakucuk A, Azim MM, Lokhat D, Ashraf GM, 2018. Essential oils: extraction techniques, pharmaceutical and therapeutic potential - a review. *Curr. Drug Metab.*, **19**: 1100-1110.
- Bajwa GA, Gul H, 2000. Some observations on insect species of *Paulownia* species at Pakistan Forest Institute Campus, Peshawar. *Pak. J. For.*, **50**: 71-80.
- Banaras S, Javaid A, Khan IH, 2020. Potential antifungal constituents of *Sonchus oleraceus* against *Macrophomina phaseolina*. *Int. J. Agric. Biol.*, **24**: 1376-1382.
- Boussaada O, Ammar S, Saidana D, Chriaa J, Chraif J, Daami M, Helal AN, Mighri Z, 2008. Chemical composition and antimicrobial activity of volatile components from capitula and aerial parts of *Rhaponticum acaule* DC growing wild in Tunisia. *Microbiol. Res.*, **163**: 87-95.
- Brehm-Stecher BF, Johnson EA, 2003. Sensitization of *Staphylococcus aureus* and *Escherichia coli* to antibiotics by the sesquiterpenoids nerolidol, farnesol, bisabolol, and apritone. *Antimicrob. Agents Chemother.*, **47**: 3357-3360.
- Cercós AP, 1982. Antimicrobial activity of the epicarp of the fruits of *Paulownia fortunei* and *Paulownia tomentosa*. *Rev. Argent. Microbiol.*, **14**: 111-114.
- Djabou N, Andreani S, Varesi L, Tomi F, Costa J, Muselli A, 2013. Analysis of the volatile fraction of *Teucriummarum* L. *Flavour Fragr. J.*, **28**: 14-24.
- Ghaffari T, Kafil HS, Asnaashari S, Farajnia S, Delazar A, Baek SC, Kim KH, 2019. Chemical composition and antimicrobial activity of essential oils from the aerial parts of *Pinuseldarica* grown in Northwestern Iran. *Molecules*, **24**: Article 3203.
- Gurib-Fakim A, Maudarbaccus N, Leach D, Doimo L, Wohlmuth H, 2002. Essential oil composition of Zingiberaceae species from Mauritius. *J. Essent. Oil Res.*, **14**: 271-273.
- He T, Vaidya BN, Perry ZD, Parajuli P, Joshe N, 2016. *Paulownia* as a medicinal tree: Traditional uses and current advances. *Eur. J. Med. Plant.*, **14**: 1-15.
- Ibrahim NA, El-Hawary SS, Mohammed MMD, Faraid MA, Nayera AM, Refaat ES, 2013. Chemical composition, antimicrobial activity of the essential oil of the flowers of *Paulownia tomentosa* (Thunb.) Steud. growing in Egypt. *J. Appl. Sci. Res.*, **9**: 3228-3232.
- Inoue Y, Shiraishi A, Hada T, Hirose K, Hamashima H, Shimada J, 2004. The antibacterial effects of terpene alcohols on *Staphylococcus aureus* and their mode of action. *FEMS Microbiol. Lett.*, **237**: 325-331.
- Javaid A, Munir R, Khan IH, Shoab A, 2020. Control of the chickpea blight, *Ascochyta rabiei*, with the weed plant, *Withania somnifera*. *Egypt. J. Biol. Pest Control*, **30**: Article 114.
- Jirovetz L, Buchbauer G, Schweiger T, Denkova Z, Slavchev A, Stoyanova A, Schmidt E, Geissler M, 2007. Chemical composition, olfactory evaluation and antimicrobial activities of

- Jasminum grandiflorum* L. absolute from India. *Nat. Prod. Commun.*, **2**: 407-412.
- Khan MF, Dwivedi AK, 2018. A review on techniques available for the extraction of essential oils from various plants. *Int. Res. J. Eng. Technol.*, **5**: 5-8.
- Khan IH, Javaid A, 2020. Antifungal activity and GC-MS analysis of *n*-butanol extract of quinoaleaves. *Bangladesh J. Bot.*, **49**: 1045-1051.
- Khatua S, Pandey A, Biswas SJ, 2016. Phytochemical evaluation and antimicrobial properties of *Trichosanthes dioica* root extract. *J. Pharmacog. Phytochem.*, **5**: 410-413.
- Krista S, Banovaca D, Tabancab N, Wedgec DE, Gochevd VK, Wannere J, Schmidta E, Jirovetz L, 2015. Antimicrobial activity of nerolidol and its derivatives against airborne microbes and further biological activities. *Nat. Prod. Commun.*, **10**: 143-148.
- Lapczynski A, Bhatia SP, Letizia CS, Api AM, 2008. Fragrance material review on nerolidol (isomer unspecified). *Food Chem. Toxicol.*, **46**: 247-250.
- Lee K-T, Koo S-J, Jung S-H, Choi J, Jung H-J, Park H-J, 2002. Structure of three new terpenoids, spiciformisins a and b, and monocyclosqualene, isolated from the herbs of *Ligularia fischeri* var. *spiciformis* and cytotoxicity. *Arch. Pharm. Res.*, **25**: 820-823.
- Lee SJ, Hans JI, Lee GS, Park MJ, Choi IG, Na KJ, Jeung EB, 2007. Antifungal effect of eugenol and nerolidol against *Microsporium gypseum* in a guinea pig mode. *Biol. Pharm. Bull.*, **30**: 184-188.
- Marrufu T, Nazzaro F, Mancini E, Fratianni, 2013. Chemical composition and biological activity of the essential oil from leaves of *Moringa oleifera* Lam. cultivated in Mozambique. *Molecules*, **18**: 10989-11000.
- Perczak A, Gwiazdowska D, Marchwińska K, Juś K, Gwiazdowski R, Waśkiewicz A, 2019. Antifungal activity of selected essential oils against *Fusarium culmorum* and *F. graminearum* and their secondary metabolites in wheat seeds. *Arch. Microbiol.*, **201**: 1085-1097.
- Pino JA, Mesa J, Muñoz Y, Martí MP, Marbot R, 2005. Volatile components from mango (*Mangifera indica* L.) cultivars. *J. Agric. Food Chem.*, **53**: 2213-2223.
- Rai AK, Singh SP, Luxmi C, Savita G, 2000. *Paulownia fortunei* - A new fibre source for pulp and paper. *Indian J. Pulp Paper Technol.*, **12**: 51-56.
- Raina VK, Verma SC, Dhawan S, Khan M, Ramesh S, Singh SC, Yadav A, 2006. Srivastava SK, 2006. Essential oil composition of *Murraya exotica* from the plains of northern India. *Flavour Fragr. J.*, **21**: 140-142.
- Reddy DN, 2019. Essential oils extracted from medicinal plants and their applications. In *Natural bio-active compounds*. Springer, Singapore. pp. 237-283.
- Rouis-Soussi LS, Ayeb-ZakhamaA, Mahjoub A, Flamin G, Jannet HB, Harzallah-Skhiri F, 2014. Chemical composition and antibacterial activity of essential oils from the Tunisian *Allium nigrum* L. *EXCLI J.*, **13**: 526-535.
- Saad SB, Ibrahim MA, Jatau ID, Shuaibu MN, 2019. Trypanostatic activity of geranylacetone: Mitigation of *Trypanosoma congolense*-associated pathological perturbations and insight into the mechanism of anaemia amelioration using *in vitro* and *in silico* models. *Exp. Parasitol.*, **201**: 49-56.
- Sarkic A, Stappen I, 2018. Essential oils and their single compounds in cosmetics-A critical review. *Cosmetics*, **5**: 11.
- Sharmeen JB, Mahomoodally FM, Zengin G, Maggi F, 2021. Essential oils as natural sources of fragrance compounds for cosmetics and cosmeceuticals. *Molecules*, **26**: Article ID 666.
- Siddiqui KM, Khan M, 1989. Introduction of *Paulownia* in Pakistan. *Pak. J. For.*, **39**: 171-176.
- Singh B, Singh JP, Kaur A, Yadav MP, 2021. Insights into the chemical composition and bioactivities of citrus peel essential oils. *Food Res. Int.*, **143**: Article 110231.
- Tariq S, Wani S, Rasool W, Shafi K, Bhat MA, Prabhakar A, Rather MA, 2019. A comprehensive review of the antibacterial, antifungal and antiviral potential of essential oils and their chemical constituents against drug-resistant microbial pathogens. *Microb. Pathog.*, **134**: Article 103580.
- Valdivieso-Ugarte M, Gomez-Llorente C, Plaza-Díaz J, Gil Á, 2019. Antimicrobial, antioxidant, and immunomodulatory properties of essential oils: A systematic review. *Nutrients*, **11**: Article 2786.
- Zhao J, Shan T, Huang Y, Liu X, Gao X, Wang M, Jiang W, Zhou L, 2009. Chemical composition and *in vitro* antimicrobial activity of the volatile oils from *Gliomastixmurorum* and *Pichia guilliermondii*, two endophytic fungi in *Paris polyphylla* var. *yunnanensis*. *Nat. Prod. Commun.*, **4**: 1491-1496.